

# Biocompatibility analysis of a novel reabsorbable alloplastic membrane composed of alginate-Capsul

Análise da biocompatibilidade de uma membrana aloplástica reabsorvível composta de alginato-capsul

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## ABSTRACT

### Objective

The aim of this study was to evaluate *in vivo* the biological response after implantation of a novel alginate-capsule membrane.

### Methods

The material was implanted into subcutaneous tissue of mice (n=15) and after 1, 3 and 9 weeks, the animals were sacrificed and biopsies analyzed with light microscopy, using the stains hematoxylin-eosin, picosirius and alcian blue pH 2.5. The parameters evaluated were: intensity and kind of inflammatory infiltrate, presence of connective tissue, foreign body reaction, vascularization and biodegradation.

### Results

1 week after implantation, the following was observed: mixed inflammatory infiltrate, absence of necrosis and beginnings of membrane fragmentation; after 3 weeks, discrete presence of multinuclear giant cells and beginnings of neovascularization; and after 9 weeks there was minor biodegradation associated with the presence of new connective tissue, and persistence of moderate inflammatory reaction observed from beginning to end of the experiment.

### Conclusion

Considering the results obtained, it is possible to conclude that the novel alginate-capsule membrane is partially reabsorbable but with low biocompatibility, requiring more tests to validate its clinical use.

**Indexing terms:** Material testing. Seaweed. Tissue engineering.

## RESUMO

### Objetivo

Avaliar *in vivo* a resposta tecidual após a implantação de uma nova membrana de alginato-capsul.

### Métodos

O material foi implantado no tecido subcutâneo de camundongos (n=15) e após 1, 3 e 9 semanas, os animais foram mortos e as biópsias analisadas à microscopia de luz, através de coloração com hematoxilina-eosina, picosirius e azul de alcian pH 2,5. Os parâmetros avaliados foram: intensidade e tipo de infiltrado inflamatório, presença de tecido conjuntivo, reação de corpo estranho, vascularização e biodegradação.

### Resultados

Após 1 semana da implantação, notou-se infiltrado inflamatório misto, ausência de necrose e início de fragmentação da membrana, em 3 semanas, observou-se presença discreta de células gigantes multinucleadas e início de neovascularização, e em 9 semanas houve pequena biodegradação associada com a presença de novo tecido conjuntivo e persistência de reação inflamatória moderada observada desde o início do experimento.

### Conclusão

Considerando os resultados obtidos concluiu-se que a nova membrana de alginato-capsul é parcialmente reabsorvível, mas com baixa biocompatibilidade, necessitando de mais testes para validar seu uso clínico.

**Termos de indexação:** Teste de materiais. Alga marinha. Engenharia tecidual.

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## INTRODUCTION

The concepts of Guided Tissue Regeneration (GTR) and Guided Bone Regeneration (GBR) were first described in the literature in 1959<sup>1-2</sup>. Both are based on four basic principles: biocompatibility, maintenance of adequate space to support the newly formed tissue, the promotion of growth of osteogenic cells and peripheral sealing to prevent growth of unwanted cellular tissue inside the defective area and the displacement of the membrane<sup>3-5</sup>. In this context, the development of new membranes that will serve these principles and permit safe, large-scale and low-cost production, has been a challenge for tissue engineering.

Alloplastic materials are mineral or synthetic in origin and, as they result from a process of industrialization, they can be supplied in adequate quantities for all needs. Due to the smaller osteoconductive capacity when compared to autogenic bone, as well as being osteoinductive and osteogenic, these materials may have different chemical formulations and associations with the strategy of improving performance<sup>6</sup>.

Sodium alginate, which derives from brown algae found in the oceans, is a linear polysaccharide formed by units of -D-mannuronic acid (M) and -L-guluronic acid (G) linked by type 1-4 glycosidic bonds, forming alginic acids. The proportion between the monomeric units is variable, as is the G or M sequence of units in the polymer chain. It is used as a thickener and emulsion stabilizer in the foods industry and as a modeler in Dentistry. It forms a reticulate gel in the presence of divalent and trivalent cations, the calcium alginate having a high incidence of G sequences (block G) being mostly used for capturing live cells and bioactive substances<sup>7</sup>. The action mechanism of the alginates is based on an ion exchange reaction between the calcium ions of the biomaterial and sodium ions present in the ooze of the wound, allowing the alginate fibers to be transformed into a soluble gel that promotes a damp microenvironment in the injury which is ideal for the process of healing<sup>8-10</sup>.

Capsul, also known as octenylsuccinate starch is obtained through the esterification of waxy starch with the acid anhydrous octenylsuccinate, which results in a hydrophobically modified starch<sup>11</sup>, capable of improving the texture of pastes or gels and the formation of film, adding hydrophobic groupings and introducing emulsifying power<sup>12</sup>.

Starch and alginate are polymers with an ability to capture molecules in technologies for the controlled release of bioactive substances<sup>13</sup>. A mixture of alginate: Capsul (3:4) produced edible film with adequate capture properties and vitamin C protection for 90 days<sup>7,14</sup>.

Therefore the aim of the present study is to analyze the biocompatibility and biodegradability of a new alginate-Capsul alloplastic membrane under the skin of mice.

## METHODS

The test material used in this study was an alloplastic membrane created from alginate (Fluka, USA) and Capsul (National Starch & Chemical, USA), in a proportion of 3:4, and then treated via immersion in a solution of calcium chloride (0.15M CaCl<sub>2</sub>), and adjusted to dimensions of 5mm x 5mm for the biocompatibility trial (Figure 1). Prior to carrying out the experiment, each material was individually packed and underwent damp heat sterilization in an autoclave (Cristófoli, Brazil), temperature=127°C, pressure=1atm, cycle time=20 minutes.

The biological characterization (biocompatibility and biodegradation of the membrane) was conducted in accordance with the procedures of ISO standard 10993-6 (International Organization for Standardization)<sup>15</sup>. This study was approved by the Ethics in Research Committee of the Fluminense Federal University (Opinion CEPA-UFF 35/08). All of the surgical procedures were carried out in the Experimental Surgery Laboratory of the Animal Facility at the Fluminense Federal University following the directives of the Brazilian College of Animal Experimentation. A total of 15 BALB/c mice were used, all 2 months old and weighing around 75g, kept in proper conditions of hygiene and overall care for the duration of the experiments.

The animals received intramuscular anesthetic using a solution composed of ketamine chlorhydrate (Dopalen®, Vetbrands, Brazil; 100 mg/kg of animal) and xylazine chlorhydrate (Anasedan®, Vetbrands, Brazil; 10 mg/kg of animal). The aforementioned dosage of anesthetic produced the desired effects (immobilization and pain suppression of the specimen) and for the period of time required to carry out the surgical activity (30 minutes). After a trichotomy and dorsal antisepsis, a 1.5cm linear incision was carried out on each animal followed by dissection of the receptor region, implantation of the biomaterial into the subcutaneous tissue (Figure 2) and suture with 5.0 nylon thread (Ethicon®, Johnson & Johnson, Brazil). After 1, 3 and 9 weeks, the animals were

put down using an anesthetic overdose and a necropsy was performed on the site of the implantation. The histological processing of the collected samples involved: fixation in 4% buffered formaldehyde (pH 7.2) for 48 hours; dehydration in increasing ethanol baths (1 bath at 70%, 80% and 90% and 3 baths at 100%, 1 hour each); diaphanization in xylene (3 baths, 30 minutes each), impregnation in liquid paraffin at 60°C (3 baths, 1 hour each); and inclusion in paraffin. Cuts 5µm thick were colored using the techniques of Hematoxylin-Eosin (HE), picosirius and alcian blue pH 2.5 and they were submitted for descriptive analysis, which evaluated the following parameters: inflammatory reaction (presence and intensity of polymorphonuclear, mononuclear and multinucleated foreign body giant cells), repair process (granulation tissue, neovascularization, fibrosis) and biodegradation (presence or fragmentation of material).

## RESULTS

### Biocompatibility

1 week - Presence observed of mixed inflammatory infiltrate (polymorpho- and mononuclear cells) of slight to moderate intensity, absence of areas of necrosis, absence of multinucleated foreign body giant cells (HE) (Figure 3A) and absence of reaction to collagen (picrosirius) (Figure 3B).

3 weeks - Presence of predominantly mononuclear inflammatory infiltrate, of slight to moderate intensity, presence of connective tissue permeating the material, discrete presence of new blood vessels and multinucleated foreign body giant cells enveloping the alginate (HE) (Figure 3C). The permeability of connective tissue is confirmed by the slight reaction to collagen (picrosirius) (Figure 3D).

9 weeks - Continuity of the mononuclear inflammatory infiltrate, of slight to moderate intensity, presence of loose connective tissue permeating the material in more significant quantities, with discrete vascularization and absence of multinucleated foreign body giant cells (HE) (Figure 3E). There is moderate reaction to collagen (picrosirius) (Figure 3F).

### Biodegradability

1 week - The alginate matrix already shows the beginnings of degradation, with more sizeable fragments, greater than 100µm in extension (HE and alcian blue pH 2.5) (Figures 4A and 4B).

9 weeks - The alginate matrix shows process of more advanced degradation, with small fragments, less than 100µm in extension (HE and alcian blue pH 2.5) (Figures 4C and 4D).

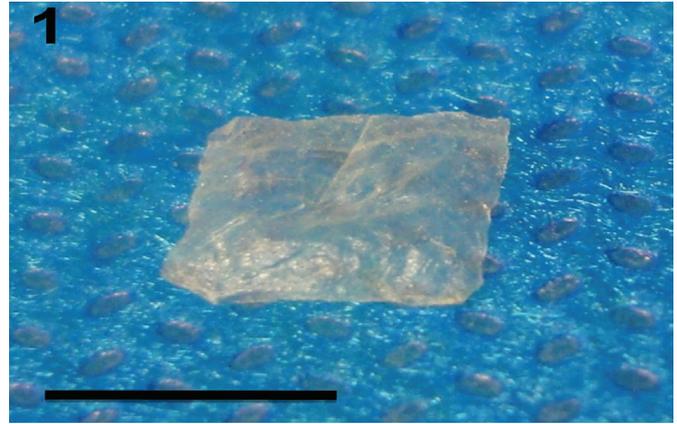


Figure 1. Alginate-Capsul membrane. The black bar represents 5mm.



Figure 2. Alginate-Capsul membrane in the mouse's subcutaneous tissue.

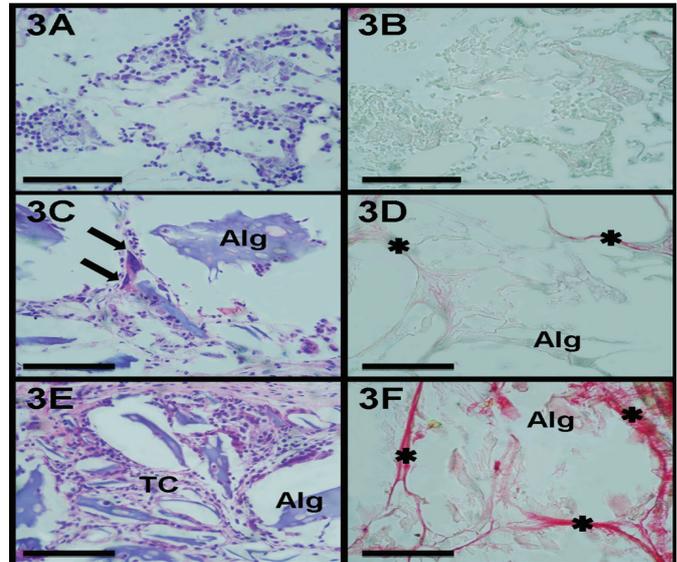
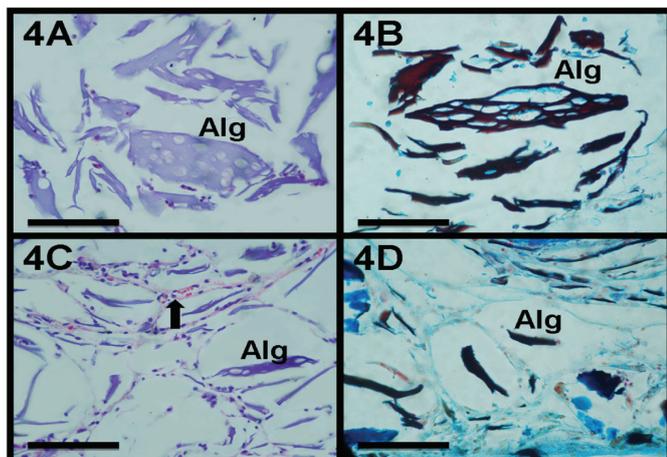


Figure 3. Biocompatibility analysis of the alginate-Capsul membrane. 1 week: mononuclear inflammatory infiltrate covering the material (A); absence of reaction to collagen (B). 3 weeks: mononuclear inflammatory infiltrate and appearance of multinucleated giant cells (black arrows), enveloping alginate (Alg) (C); minor reaction to collagen (D). 9 weeks: persisting mononuclear inflammatory infiltrate and abundant loose connective tissue permeating the material (E); moderate reaction to collagen (F). A, C, E: Hematoxylin-Eosin; B, D, F: Picrosirius. The black bar represents 100µm.



**Figure 4.** Analysis of biodegradability of alginate-Capsul membrane. 1 week: presence of larger fragments of alginate (A-B). 9 weeks: presence of smaller fragments of alginate (C-D). A, C: Hematoxylin-Eosin; B, D: Alcian blue pH 2.5. The black bar represents 100µm.

## DISCUSSION

Alginate is a natural polysaccharide whose biological behavior is being increasingly analyzed through tissue engineering. For many years it has been recognized as a biomaterial capable of causing inflammatory reaction. By itself, the surgical procedure for implantation of a material is capable of initiating an inflammatory response, however when in contact with the tissue, alginate induces macrophage activation<sup>16</sup>.

The characterization of the chemical structure of the alginate is still insufficient, however it is known that it presents lipopolysaccharide, which is a powerful activator of innate immune response<sup>17</sup>. An important regulator, of both innate immune response and adaptive immune response, recognized as a nuclear transcription factor (NF- $\kappa$ B), it is activated when the organism comes into contact with alginate, stimulating macrophages to produce proinflammatory cytokines such as IL-1, IL-6, IL-12 and TNF- $\alpha$ <sup>18-19</sup>.

The in vivo implantation of unpurified alginate is shown to be favorable, and is capable of stimulating cell proliferation<sup>20</sup>. In smaller bone defects in rabbits, an alginate membrane compared with a collagen showed double the bone neoformation (in 8 weeks) and degradation that was twice as fast (18 to 28 days and 28 to 56 days, respectively) in mandibular defects ( $\Theta=5$ mm)<sup>21</sup>, and a greater presence of osteoinductive factors (VEGF, TGF- $\beta$  and BMP) in defects of the tibia ( $\Theta=5$ mm)<sup>22</sup>. Associating alginate with other components, such as hydroxyapatite, may have a positive

influence on bone and vascular neoformation where there is extensive bone loss, as observed in skull defects in rats ( $\Theta=8$ mm) in later periods (120 days)<sup>20</sup>.

Control over the degradability of the biomaterial is frequently a key factor in tissue replacement, since it will serve as the skeleton for growth and the integration of the new tissue<sup>23</sup>. The quantity and quality of new tissue to be formed are directly related to the speed of alginate degradation. Accordingly, gels with very fast degradation (100% volume loss in 5 days) prevent the maintenance of the tissue skeleton, while gels with moderate to slow degradation (less than 20% in 4 weeks) favor the proliferation of osteoblasts and stimulate the production of mineralized matrix<sup>24</sup>.

## CONCLUSION

According to the results presented, the alginate-Capsul membrane presents favorable biodegradation and is capable of acting as a skeleton for cell proliferation and tissue neoformation. However, studies should be performed with a view to minimizing or controlling the intensity of the chronic inflammatory process triggered by its implantation. Modifications in its processing and purification, in addition to more pre-clinical studies, may contribute to the development of the material.

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## Collaborators

C JARDELINO contributed with experimental surgery, histological processing, histological colorations, data interpretation and the composition of the article. II CASTRO-SILVA contributed with the histopathological analyses and the composition of the article. CPG MACHADO contributed

with experimental surgery and the composition of the article. MH ROCHA-LEÃO, AM ROSSI and SRA SANTOS contributed to the development and production of test material and the

composition of the article. JM GRANJEIRO contributed to the development of the test material, final interpretation of the data and the composition of the article.

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